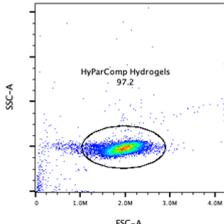
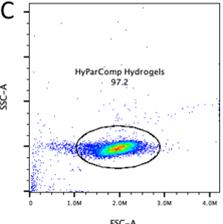
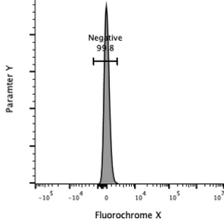
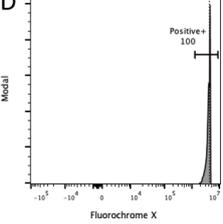


1. Technical Data Sheet

<p>Summary</p>	<p>HyParComp™ compensation controls are state-of-the-art cell mimics that capture multiple antibody host species (mouse anti-human, mouse, rat, and hamster), and mimic the fluorescence spectra of stained cells.</p>
<p>Application</p>	<p>HyParComp™ are intended as compensation controls to match the single staining performance of real cells. Staining the cell mimics yields a positive fluorescence histogram that will aid in resolving the performance of the fluorophore; it will also serve as the basis for positive signal of a given fluorophore for compensation and/or spectral unmixing.</p> <p>Note: HyParComp™ performance has been verified and validated on analytical flow cytometers and not on cell sorters.</p> <p>For Research Use Only. Not for use in diagnostic or therapeutic procedures.</p>
<p>Materials</p>	<p>HyParComp™ are cell mimics that are suspended in aqueous solution and are packaged in a convenient dropper bottle. Each drop contains approximately 1×10^5 beads.</p>
<p>Handling and Safety</p>	<p>No special handling or safety precautions are necessary. See the Safety Data Sheet (SDS) at www.slingshotbio.com.</p>
<p>Instructions for Use</p>	<ol style="list-style-type: none"> 1. Unpack and vortex both vials on high for 2 - 3 seconds to resuspend cell mimics. 2. Add 1 drop of the negative cell mimics into the bottom of the test tube or well of a plate for the unstained negative control. (1 drop contains approximately 1×10^5 cell mimics). 3. Add 1 drop of the positive capture cell mimics into the bottom of the test tube or well of a plate for each fluorophore you will have in the experiment. (1 drop contains approximately 1×10^5 cell mimics). 4. Add your pre-titrated antibody directly to the mimics and vortex. <p>Note: It is recommended to pre-determine the appropriate titer of the antibody that works best for the application. DO NOT add antibody to the unstained tube.</p> <ol style="list-style-type: none"> 5. Incubate at room temperature for 15 - 30 minutes, protected from light. <p>Note: Particles should be treated the same way as cells to be analyzed, i.e. all fixation and permeabilization steps used on cells should be applied to the particles. <i>Do not add Brilliant Violet Staining Buffer or SuperBright Staining buffer to single stained controls.</i></p>

	<p>6. Add 2 ml of 1X PBS containing 1% BSA (Bovine Serum Albumin) to the tube.</p> <p>Note: Staining buffer containing BSA or FBS (Fetal Bovine Serum) can also be used for washing.</p> <p>7. Centrifuge the tube for 5 minutes at 600 g and immediately aspirate the supernatant to minimize the cell mimic loss, being careful not to disturb the cell mimic pellet.</p> <p>8. Resuspend the cell mimic pellet in 1X PBS at preferred volume and vortex.</p> <p>Note: Protect the samples from light and analyze the samples as soon as possible.</p> <p>9. View and acquire the HyParComp cell mimics on Forward and Side Scatter parameters (FSC-A and SSC-A) using the same instrument settings as leukocytes.</p>
<p>Storage</p>	<p>HyParComp™ should be stored at 2 - 8 °C once the product is received.</p>
<p>Expiration</p>	<p>One year from the date of manufacturing</p>
<p>QC Data</p>	<p style="text-align: center;">HyParComp Figure 1 (A, B, C, D)</p> <div style="display: flex; justify-content: space-around;"> <div style="text-align: center;"> <p>A</p>  </div> <div style="text-align: center;"> <p>C</p>  </div> </div> <div style="display: flex; justify-content: space-around; margin-top: 20px;"> <div style="text-align: center;"> <p>B</p>  </div> <div style="text-align: center;"> <p>D</p>  </div> </div> <p style="margin-top: 20px;">Figure 1. (A) Gate the HyParComp population from the negative sample. (B) Place a gate on the negative histogram for the fluorophore of interest from the negative sample. (C) Gate the HyParComp population from a positive single-stain control sample. (D) Place a gate on the positive histogram for the fluorophore of interest from the positive single-stain control sample.</p>